



A Preliminary Checklist of Vascular Plants Endemic to the Chapada Diamantina, Bahia, Brazil, with Comments on Their Extinction Threats Status

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ABSTRACT – The Chapada Diamantina is the northern extension of the Espinhaço Mountain Range and comprises diverse and complex vegetation types. It is also characterized by a high degree of plant endemism, especially in the “campos rupestres” sites. We present a preliminary checklist of the endemic vascular plants of Chapada Diamantina within the concept of the “Chapada Diamantina Complex” ecoregion. Regional floras and inventories were searched for citations of endemism. The checklist includes 459 endemic plant species from the Chapada Diamantina, distributed among 48 botanic families and 156 genera. Eight genera comprise more than 10 endemic species each. *Microlicia* was the most diverse genus, with 38 species. This checklist has a preliminary character, as very few groups have been intensively studied over the entire range, and the flora of several localities is still poorly known. As the first effort to synthesize the literature concerning endemism in the Chapada Diamantina, this checklist represents an initial approximation of the wide diversity of species found there. The available data indicates that diversity is largely restricted to just a few groups. A total of 285 species were evaluated on their threats of extinction. Of these, 277 species have some degree of threat, 40 (14.44%) of them being CR. The factor influencing the extinction threat are related mainly to the narrow area of occurrence of many species. Deforestation and burning in the vegetation are other risks. On the other hand, several endemic species not evaluated are relatively common in the Chapada Diamantina and aren't on threat of extinction.

Keywords: Chapada Diamantina; endemics; rupestrian grasslands.

Lista Preliminar de Plantas Vasculares Endêmicas da Chapada Diamantina, Bahia, Brasil, com Comentários sobre seus Status de Ameaça de Extinção

RESUMO – A Chapada Diamantina é a extensão norte da cadeia do Espinhaço e caracteriza-se por vegetações diversas e complexas. Também é caracterizada por muito endemismo das plantas, especialmente nos campos rupestres. Neste artigo é apresentada uma lista preliminar das plantas vasculares endêmicas de Chapada Diamantina. O conceito de Chapada Diamantina adotado aqui é a ecorregião “Complexo da Chapada Diamantina”. A literatura botânica sobre floras regionais e inventários botânicos locais foi pesquisada por citações do endemismo. A lista retornou 459 espécies de plantas endêmicas da Chapada Diamantina, distribuídas entre 48 famílias botânicas e 156 gêneros. Oito gêneros compreenderam mais de 10 espécies endêmicas cada. *Microlicia* foi o gênero mais diversificado, com 38 espécies. Essa lista tem um caráter preliminar, pois poucos grupos na Chapada Diamantina foram intensamente estudados em toda a área, e a flora de várias localidades ainda é pouco conhecida. Como o primeiro esforço para sintetizar a literatura sobre o endemismo na Chapada Diamantina, a lista apresenta uma aproximação inicial da grande diversidade de espécies encontradas. Os dados disponíveis indicam, no entanto, que a diversidade é restrita principalmente

a apenas alguns grupos. Um total de 285 espécies foram avaliadas em suas ameaças de extinção. Dessas, 277 espécies apresentam algum grau de ameaça, sendo 40 (14,44%) delas Criticamente em Perigo. Os fatores que influenciam a ameaça de extinção estão relacionados principalmente à pequena área de ocorrência de muitas espécies. Desmatamento e queimadas na vegetação são outros riscos. Por outro lado, várias espécies endêmicas não avaliadas são relativamente comuns na Chapada Diamantina e não estão ameaçadas de extinção.

Palavras-chave: Campos rupestres; Chapada Diamantina; endemismo.

Lista Preliminar de Plantas Vasculares Endêmicas de Chapada Diamantina, Bahía, Brasil, con Comentarios sobre su Estado de Peligro de Extinción

RESUMEN – Chapada Diamantina es la extensión norte de la sierra de Espinhaço y se caracteriza por una vegetación diversa y compleja. También se caracteriza por un alto grado de endemismo vegetal, especialmente en campos rupestres. Este artículo presenta una lista preliminar de plantas vasculares endémicas de Chapada Diamantina. El concepto de Chapada Diamantina adoptado aquí es la ecorregión “Complejo da Chapada Diamantina”. Se buscó en la literatura botánica sobre floras regionales e inventarios botánicos locales citas de endemismo. La lista arrojó 459 especies de plantas endémicas de Chapada Diamantina, distribuidas en 48 familias botánicas y 156 géneros. Ocho géneros comprendían más de 10 especies endémicas cada uno. Microlicia fue el género más diverso, con 38 especies. Esta lista tiene un carácter preliminar ya que pocos grupos en Chapada Diamantina han sido estudiados intensamente en toda el área y la flora de varios lugares aún es poco conocida. Como primer esfuerzo de síntesis de la literatura sobre endemismo en la Chapada Diamantina, esta lista presenta una primera aproximación a la gran diversidad de especies encontradas. Los datos disponibles indican, sin embargo, que la diversidad se restringe principalmente a unos pocos grupos. Un total de 285 especies fueron evaluadas en sus amenazas de extinción. De estas, 277 especies presentan algún grado de amenaza, siendo 40 (14.44%) de ellas CR. Los factores que influyen en la amenaza de extinción están relacionados principalmente con la estrecha área de presencia de muchas especies. La deforestación y la quema de la vegetación son otros riesgos. Por otro lado, varias especies endémicas no evaluadas son relativamente comunes en la Chapada Diamantina y no están en peligro de extinción.

Palabras clave: Campos rocosos; Chapada Diamantina; endemismo.

Introduction

The Chapada Diamantina Ecoregion (Vellozo et al., 2002) represents the northern portion of the Espinhaço Range, located in the center of Bahia State, in northeastern Brazil. It is characterized by a diverse and complex vegetation, with elements from three different Brazilian biomes: Cerrado (Brazilian savannah), Atlantic Forest (mainly seasonal forests), and Caatinga (composed mainly of xerophytic formations), as well as several ecotones (Velloso et al., 2002; Zappi et al., 2003; Giuliatti et al., 2004; Conceição et al., 2007). The highest elevations of the region are occupied by “Campos Rupestres” (Rupestrian Grasslands), which are recognized as biodiversity hotspots, with high degrees of plant

species richness, endemism, and unique species compositions (Conceição & Pirani, 2016). This area is also cited as having high faunal endemism (Heyer, 1999; Napoli & Juncá, 2006; Rodrigues et al., 2006; 2009; Napoli et al., 2011; Pombal Jr. et al., 2012; Teixeira Jr. et al., 2012; Fernandes & Handam, 2014).

Conceição et al. (2016) recorded 165 species endemic to the Espinhaço Range in 785 plots of campos rupestres sampled within 39 communities, which represented 32.7% of all of the species in their study. Several other studies have emphasized the high numbers of endemic species in the Campos Rupestres of the Chapada Diamantina (Guedes & Orgue, 1998; Stannard, 1995; Zappi et al., 2003). Many endemic species have likewise

been described from other formations in the Chapada Diamantina, such as seasonal forests (Fiaschi, 2005; Santo et al., 2012) and Caatinga (Vasconcelos et al., 2016; Silva-Castro, 2017).

Lists of endemic species can be used to justify the creation of protected areas and conservation of natural environments (Hazevoet, 1996; Grill et al., 2002). Forzza et al. (2010) reported that 56% of the Brazilian flora is restricted only to that country. Giulietti et al. (2002) listed 318 species endemic to the Caatinga Biome. In spite these references and lists of endemics of other taxonomic groups (Martinelli et al., 2008, Versieux et al., 2008, Conceição et al., 2015) or published accounts of endemic species in regional floras (Romero & Nakajima, 1999; Siqueira Filho et al., 2012), checklists of the endemics of specific Brazilian regions are uncommon. Thomas et al. (1998) listed 394 taxa (including species, subspecies, and varieties) in their checklist of the endemic taxa of two forest areas in southern Bahia and Espírito Santo State.

This work presents a preliminary checklist of endemic vascular plants of the Chapada Diamantina. Discussions of taxonomic aspects, conservation status, and other relevant topics of the species listed are also provided.

Materials and Methods

The concept of endemism is useful for quantifying the biological uniqueness of an area. Peterson & Watson (1994) argued, however, that it is important differentiate between a geographic definition (the restricted range of distribution of a given taxon) and a regional distribution approach (endemism). In this work, we consider those species restricted to the Chapada Diamantina region (as defined below) as endemics, although that view encompasses several distribution ranges within that region.

Velloso et al. (2002) proposed several ecoregions for the Caatinga biome, including the Chapada Diamantina Complex, which is roughly equivalent to the relief concept of IBGE (2006), and includes considerations of soils, climate, vegetation, geomorphology, and geology. We

adopted that concept as the delimitation of the Chapada Diamantina (Figure 1). As mentioned previously, that delimitation includes xerophytic formations, grasslands and savannah-like formations, Rupestrian Grasslands, seasonal forests, and small patches of evergreen forest, as well as the ecotones between those formations (Velloso et al., 2002).

We searched the botanical literature for regional floras and inventories looking for citations of endemism in the Chapada Diamantina. The extensive work of Giulietti et al. (2009), evaluating the rare species of Brazilian flora, as well as the Red Book of the Brazilian Flora (Martinelli & Moraes, 2013), furnished some basic information for several groups. We also looked for species descriptions and revisions of genera or families where the geographical distributions of their species were cited. We listed only species names (not considering subspecies or varieties). The species names were checked for synonyms using the taxonomic literature available for each family. The concepts adopted for defining botanical families are based on APG IV (2016) and PPG I (2016).

All data obtained were listed on a matrix containing the species name and geographic information. We used the Specieslink (www.splink.cria.org.br) as a database to review the distribution matrix and to evaluate any eventual errors concerning the citations of species as endemic to the Chapada Diamantina. The species descriptions available on the site www.ipni.org were used, in some cases, as an additional data source if the species was not listed on the Specieslink.

Data about the evaluation of extinction risk for the endemic plant species to the Chapada Diamantina were obtained in the “Lista Nacional Oficial de Espécies da Flora Ameaçadas de Extinção” (Official National List of Endangered Species of Flora; MMA, 2014; National list, henceforth) and in the “Lista Oficial das Espécies Endêmicas da Flora Ameaçadas de Extinção do Estado da Bahia” (Official List of Endemic Flora Species Threatened by Extinction from the State of Bahia; Bahia, 2017; Bahia list, henceforth). Following these official lists, the species were

classified in categories based on IUCN (2012), according to Table 1. Some species described in recent years were not evaluated in National or

Bahia list but their threat statuses were appointed by their authors, and we consider these evaluations on the list in the Table 2.

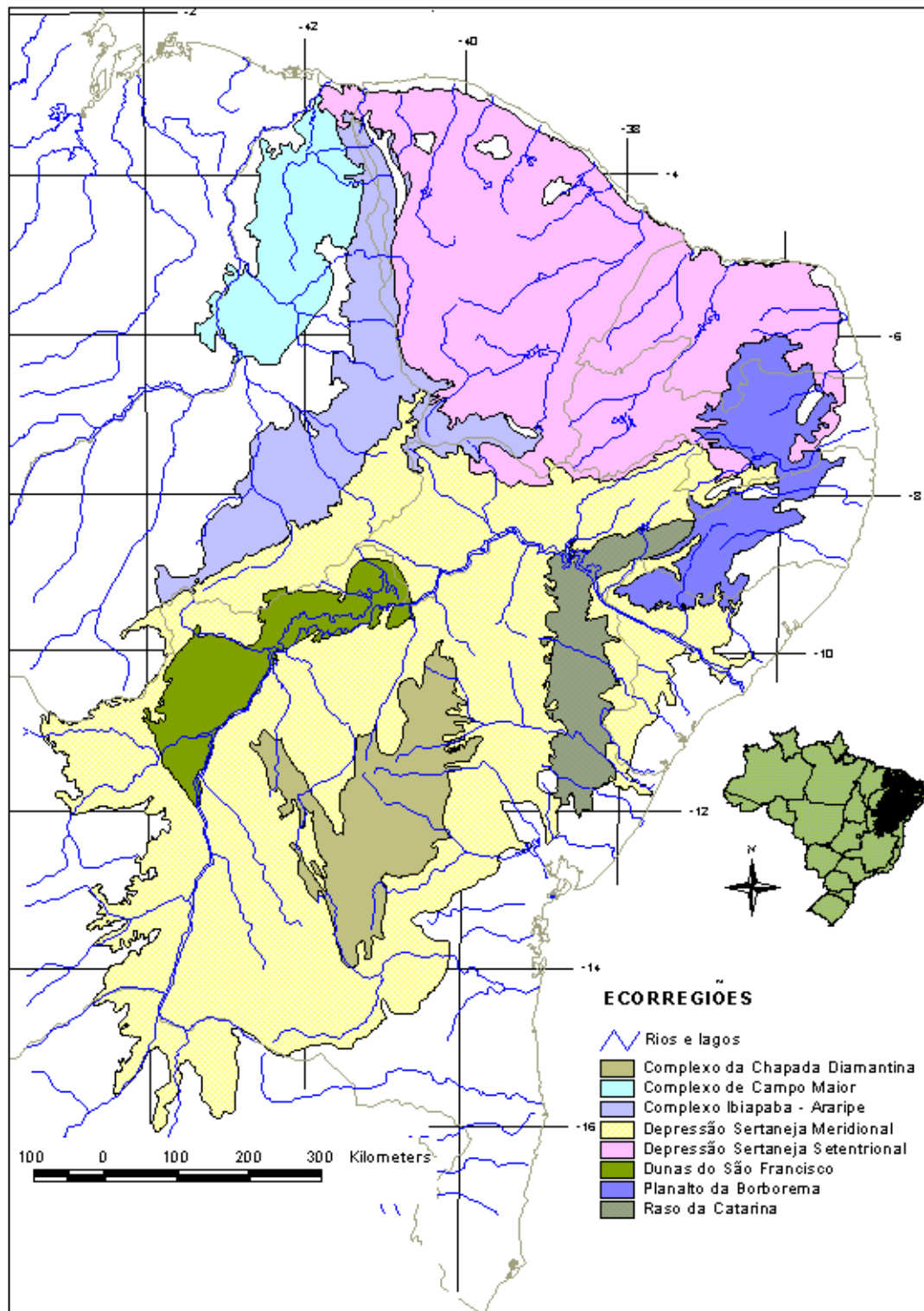


Figure 1 – The ecoregions of the Caatinga Biome as defined by Velloso et al. (2002), including the Chapada Diamantina ecoregion (“Complexo da Chapada Diamantina”). This concept includes considerations of relief, soils, climate, vegetation, geomorphology, and geology, and is used here as the basis to define the plant endemism in the Chapada Diamantina.

Table 1 – Extinction risks categories adopted in the lists of endangered species considered in this work, according to IUCN (2012).

Acronym	Category
CR	Critically endangered
EN	Endangered
VU	Vulnerable
NT	Near threatened
LC	Least concern
DD	Data deficient
NE	Not evaluated

Results

A total of 459 endemic plant species were identified for the Chapada Diamantina (Table 2), distributed among 48 botanic families and 156 genera. Asteraceae was the family with the greatest number of species (81), followed by Melastomataceae (76), Fabaceae (51), Lamiaceae (29), Bromeliaceae (24), Eriocaulaceae (19), Verbenaceae (17), Orchidaceae (16), Passifloraceae (15), and Poaceae (14). Those nine families together comprised 73.86% of the endemic species. All the other families were represented by less than 10 species; and 14 had only one endemic species. Only one endemic species belonged to a fern family (*Doryopteris trilobata* J. Prado, Pteridaceae), with all the others being angiosperms.

Seven genera showed more than 10 endemic species each. *Microlicia* was the most

diverse genus, with 37 species, followed by *Calliandra* (35), *Marcetia* (18), *Stachytarpheta* (13), *Eriope* (12), *Sincoraea* (11), and *Paepalanthus* (11). Those seven genera together comprised 29.58% of the endemic species. Twelve genera were considered endemic to the Chapada Diamantina: *Arrojadocharis*, *Bishopiella*, *Catolesia*, *Lapidia*, *Scherya*, *Semiria*, and *Stylotrichium* (all Asteraceae); *Adamantina* and *Thelyschista* (Orchidaceae); *Pseudiris* (Iridaceae); *Rayleya* (Malvaceae); and *Rupestrea* (Melastomataceae).

To date, 285 (61.77%) endemic species from Chapada Diamantina Ecoregion have their status of threat evaluated (Figure 2). Only two was considered LC, while six others were DD. All the remainder 277 species have some degree of threat, 40 (14.44%) of them being CR.

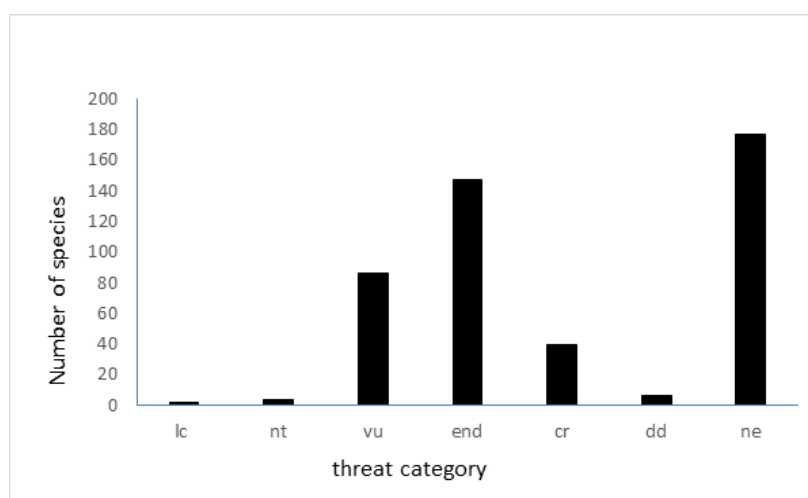


Figure 2 – Number of species endemics from Chapada Diamantina per categories of extinction risk. Acronyms according to Tab. 1.

Eleven species included in our list has nor in the National list neither in the Bahia list but were evaluated by their authors when they were described. Five of them were considered EN by their authors, four were CR, one vulnerable VU and one DD.

The National list and the Bahia list have 274 species when combined. Of these, 136 were on the Bahia list but not in the National list. On the same way, five species have different status in the Bahia list in relation to the categorization in the National list, all of them in a greater degree of threat. On the other hand, 43 species cited on the National list are not in the Bahia list.

Discussion

This checklist has a preliminary character as very few groups have been intensively studied in the whole extent of the Chapada Diamantina, and the flora of several localities is still poorly known. Intensive sampling efforts in some localities, such as the Pico das Almas (Rio de Contas), Catolés (Abaíra), Serra da Chapadinha (Lençóis), and the Pai Inácio Mountain (Palmeiras), have resulted in floras or checklists (Stannard, 1995; Guedes & Orgue, 1998; Zappi et al., 2003). Several new species have been described because of those efforts (Harley, 1988, 1992; Giulietti & Parra, 1995; Woodgyer & Zappi, 2009), and new species are still regularly described from several different areas (e.g., Giulietti & Silva, 2016; Siniscalchi et al., 2016; Vasconcelos et al., 2016; Roque et al., 2017; Silva-Castro, 2017). Some species previously listed as restricted from Chapada Diamantina, on the other hand, have been found in other localities (Monteiro et al., 2012).

As a first effort to synthesize published studies concerning endemic species in the Chapada Diamantina, this checklist presents an idea of the wide diversity of species restricted to that area. The available data indicates that the observed diversity is largely restricted to just a few groups, with ten families (17.85% of the total) retaining more than two thirds of the endemic species. Similarly, the 10 most diverse genera (5.78%) include almost one third of the endemic species.

Our knowledge of the flora from Chapada Diamantina, including its endemic species, has been greatly improved by the intensive works of

several researchers, especially A. M. Giulietti and R. N. Harley. The former described 11 of the 26 species of Eriocaulaceae listed here (Giulietti & Parra, 1995; Giulietti & Miranda, 2009; Giulietti & Silva, 2016; Pereira et al., 2016) and some species of other families (Gil et al., 2008; Campos et al., 2010). Dr. Harley described 26 of the 33 Lamiaceae species restricted to the Chapada Diamantina (Harley 1988, 1992; Harley & Walsingham, 2014), as well as species from several other groups (Zappi & Harley, 1992; Taylor et al., 2000, Harley & Giulietti, 2005).

The genera of Asteraceae restricted to Chapada Diamantina all belong to the subtribe Gyptidiane (Rivera et al., 2016; Roque et al., 2017). Rivera et al. (2016) noted that the generic concepts of this subtribe will need to be examined in more detail to precisely define their limits, and current generic concepts will probably be shifted to accommodate taxonomic classifications in response to the phylogenetic relationships identified in their analyses. Similarly, Roque et al. (2017) described the genus *Lapidia* to accommodate a shrub species that is phylogenetically basal to a clade including the genera *Catolesia*, *Morithamnus*, and *Bahianthus*. Rivera et al. (2016) cited *Agrianthus* as an endemic genus, but all of the records of *Agrianthus campestris* Mart. ex DC. are from localities in Minas Gerais State, as are most records of *A. myrtooides* Mattf. (Specieslink 2017). The remaining genera restricted to the Chapada Diamantina (*Adamantina*, *Thelyschista*, *Pseudiris*, *Rayleya*, and *Rupestrea*) are phylogenetically basal in their groups, and their actual ranges of distribution could represent remnants of wider historical distributions (van den Berg & Chase, 2004; Gil et al., 2008; Whitlock & Hale, 2011; Goldenberg et al., 2015).

Santos & Silva (2005) recorded 123 species of Melastomataceae belonging to 55 genera in the municipality of Rio de Contas, at the southwestern end of the Chapada Diamantina, with 59 endemics. Other workers have described other new endemic species of this family, mainly from Mucugê (Almeda & Martins, 2012; Freitas et al., 2012; Freitas & van den Berg, 2016; Goldenberg et al., 2015).

Fabaceae is a very diverse family in the Chapada Diamantina, mainly in terms of the genus *Calliandra*. The taxonomy of this genus in the Chapada Diamantina was reviewed by Souza (2001), who reported 36 endemic species



and pointed out the importance of Chapada Diamantina to the genus, as approximately 25% of all known *Calliandra* species are restricted to that area. Souza et al. (2013) showed that all but one endemic species belonged to a single infrageneric taxon (Sect. *Monticola*); only *C. pilgerana* Harms belongs to Sect. *Androcaullis*. The authors attributed this unbalanced diversity of the infrageneric groups of *Calliandra* to their relatively recent and rapid adaptive radiation into the Campos Rupestres, a pattern likewise observed in other groups (Souza et al., 2013).

Versieux et al. (2008) prepared a list of the Bromeliaceae from the Cadeia do Espinhaço mountain chain, which includes the Chapada Diamantina, and recorded 34 species endemic to the Chapada; one species had no specific epithet (“*Dyckia* sp 5”) in their check list and probably represents an undescribed species (which was accordingly excluded from the present list). Other bromeliad species were excluded because they are not endemic to the Chapada Diamantina (based on Specieslink 2017), including *Aechmea bahiana* L. B. Sm., *Cryptanthus warren-loosei* Leme, *Dyckia nervata* Rauh., and *Hohenbergia vestita* L. B. Smith. We list here 25 bromeliads as endemics.

A previous study focusing on the family Orchidaceae from the Chapada Diamantina listed 15 species and two genera as being endemic (van den Berg & Azevedo, 2005). Various other studies focusing on the floristic compositions of Orchidaceae in several localities cited endemic taxa (Azevedo & van den Berg, 2007; Bastos & van den Berg, 2012; Vieira et al., 2014). Additional studies, however, have shown that some of those species have relatively wide distributions and are not endemics, such as *Encyclia alboxanthina* Fowlie (Monteiro et al., 2012). Carvalho et al. (2016) discussed the taxonomic relationships between *Gomeza spilopectera* (Lindley) M. W. Chase & N. H. Williams and *Gomeza sincorana* (Campacci & Cath.) M. W. Chase & N. H. Williams and concluded that the latter is an autonomous taxon restricted to Chapada Diamantina. We recognized here 16 orchid species and two genera (*Adamantina* and *Thelyschista*) as being endemic.

When the “classic” concept of Passifloraceae is considered, this family has just one endemic taxon in the Chapada Diamantina, *Passiflora mucugeana* T. S. Nunes & L. P. Queiroz (Nunes & Queiroz, 2001; 2007). The extended concept of that family adopted in APG IV (2016) based

on phylogenetic studies (Chase et al., 2002; Tokuoka, 2012), includes the genera *Turnera* and *Piriqueta* (formerly Turneraceae), resulting in 15 species belonging to Passifloraceae being considered endemic in the present study. In the same way, *Rayleya bahiensis* is now included in Malvaceae (formerly Sterculiaceae, see Whitlock & Hale, 2011), and the genera *Barjonia*, *Hemipogon*, *Cynanchum*, *Matelea*, and *Metastelma* are included in Apocynaceae (formerly Asclepiadaceae, see Potgieter & Albert, 2001).

Species lists are important elements for conservation efforts (Forzza et al., 2010; Grill et al., 2002). Hazevoet (1996) pointed out the value of varietal or subspecific statuses of birds for island lists and considered the loss of some genetic pounds may occur when these subspecific taxa are not properly protected. We did not consider any varietal or subspecific statuses in the present study because there is no a pattern to define these status and their significance on plants (Hamilton & Reichard, 1992). Pereira et al. (2007) analyzed the morphological and genetic variability of *Comanthera mucugensis* (Giul.) Parra & Giul. (= *Syngonanthus mucugensis* Giul.) in two isolated populations, one in Mucugê in the Serra do Sincorá Range (the eastern group) and the other in Rio de Contas and Catolés near Abaíra (the western group). Those authors concluded that the level of genetic identity between those populations was less than mean values commonly found for conspecific populations, and closer to values recorded for populations of closely related congeneric species – although their morphological differences were restricted only to the simple sizes of the morphological characters analyzed; they therefore proposed a subspecific status to the western population (*C. mucugensis* ssp. *riocontensis* A.C.S. Pereira & Giul.). This was a unique study, defining the taxonomic statuses of the infraspecific taxa within an endemic species of the Chapada Diamantina. In spite those considerations, we refer to *C. mucugensis* here as just a single taxon in this checklist, following CNCFlora (2012), in which *C. mucugensis* is listed as an endangered species (EN).

Several endemic species are restricted to narrow areas. This aspect is one of possible explanations to the fact that 59.82% of all endemic species are on some degree of threat (97,19% of evaluated species). Obviously, the efforts to



evaluate the species are concentrated on those are recognized as endangered. For example, *Cattleya elongata* Barb. Rodr. (Orchidaceae) is a very common species in the Chapada Diamantina, being one endemic species with no risk of extinction (Van den Berg, 2020), and it was not included on the efforts to elaborate the National and the Bahia lists. Other aspect is existence of specialists on the taxonomic groups where are endemic species. Velloziaceae has eight endemic species in the Chapada Diamantina, but only the two species of *Barbacenia* were evaluated. All the remainder species, from the genus *Vellozia*, were not evaluated, and there are no specialists in this genus acting in this region. For other groups, as Fabaceae and Asteraceae, there are some research groups from universities in the Bahia State developed intensive studies on these taxa, and these groups are well represented in the effort to evaluate the threat of extinction. When the National and the Bahia lists are combined, Asteraceae has 56 species evaluated and Fabaceae has 22.

There are no systematizations of the impacts influencing the extinction risks for the endemic species in the Chapada Diamantina. Giulietti et al. (2004) cited the deforestation and the occurrence of fire in the vegetation as one of the most intense impacts for the Caatinga biome, including the Chapada Diamantina. Conceição et al. (2016) cited presence of invasive species has other risk for species in mountain outcrops, also including the area considered in this work. On the other hand, Conceição (2018) showed that *Vellozia pyrantha*, a fire dependent species, is threatened by the policy of fire suppression adopted in the Chapada Diamantina National Park.

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Table 2 – The preliminary checklist of vascular plants endemic to the Chapada Diamantina, Bahia, Brazil, including threat categories according to National and Bahia Lists. Acronyms according to Table 1. Asterisks (*) indicate species whose extinction risks were indicated in the articles that described them.

Family	Specific epithet	Threat Category		
		National	Bahia	Combined
Amaranthaceae	<i>Gomphrena chrestoides</i> C.C.Towns.	NT	VU	VU
	<i>Gomphrena nigricans</i> Mart.	CR	CR	CR
Apocynaceae	<i>Metastelma giuliettianum</i> Fontella	EN	EN	EM
	<i>Metastelma harleyi</i> Fontella	EN	EN	EN
	<i>Minaria harleyi</i> (Fontella & Marquete) Rapini & U.C.S.Silva	EN	NE	EN
	<i>Minaria volubilis</i> Rapini & U.C.S.Silva	EN	NE	EN
	<i>Monsanima morrenioides</i> (Goyder) Liede & Meve	CR	CR	CR
Aquifoliaceae	<i>Ilex auricula</i> S.Andrews	CR	CR	CR
	<i>Ilex mucugensis</i> Groppo	EN	EN	EN
Araceae	<i>Philodendron pachyphyllum</i> K.Krause	EN	EN	EN
Araliaceae	<i>Dendropanax geniculatus</i> Fiaschi	NE	VU	VU
Arecaceae	<i>Attalea seabrensis</i> Glassman.	NE	NE	NE
	<i>Syagrus harleyi</i> Glassman.	NE	NE	NE
Asteraceae	<i>Acritopappus catolesensis</i> D.J.N. Hind & Bautista.	VU	VU	VU
	<i>Acritopappus connatifolius</i> (Soar.Nunes) R.M.King & H.Rob.	EN	EN	EN
	<i>Acritopappus harleyi</i> R.M.King & H.Rob.	NE	VU	VU
	<i>Acritopappus jacobaeus</i> Bautista, Rodr.Oubiña & S.Ortiz	NE	NE	NE
	<i>Acritopappus morii</i> R.M.King & H.Rob.	NE	EN	EN
	<i>Acritopappus pintoii</i> D.J.N.Hind & Bautista.	CR	CR	CR
	<i>Acritopappus prunifolius</i> R.M.King & H.Rob.	NE	EN	EN
	<i>Acritopappus stenophyllus</i> Bautista, S.Ortiz & Rodr.Oubiña.	NE	NE	NE
	<i>Agrianthus almasensis</i> D.J.N.Hind.	END	END	END
	<i>Agrianthus carvalhoi</i> D.J.N.Hind	NE	VU	VU
	<i>Agrianthus corymbosus</i> DC.	NE	NE	NE
	<i>Agrianthus empretrifolius</i> Mart.	NE	NE	NE
	<i>Agrianthus giuliettiae</i> D.J.N.Hind	EN	EN	EN
	<i>Agrianthus luetzelburgii</i> Mattf.	NE	VU	VU
	<i>Agrianthus microlicioides</i> Mattf.	NE	EN	EN
	<i>Agrianthus myrtooides</i> Mattf.	NE	EN	EN
	<i>Agrianthus pungens</i> Mattf.	NE	EN	EN
	<i>Arrojadocharis praxeloides</i> (Mattf.) Mattf.	NE	EN	EN
	<i>Arrojadocharis santosii</i> R.M.King & H.Rob.	NE	NE	NE
	<i>Aspilia almasensis</i> D.J.N.Hind.	VU	NE	VU
	<i>Baccharis orbiculata</i> Deble & A.S.Oliveira	NE	EN	EN
	<i>Bishopiella elegans</i> R.M.King & H.Rob.	NE	EN	EN
	<i>Calea morii</i> H.Rob.	NE	NE	NE

	<i>Catolesia huperzioides</i> Roque, H.Robinson & A.A.Conceição	CR	NE	CR
	<i>Catolesia mentiens</i> D.J.N.Hind	CR	CR	CR
	<i>Catolesia monocephala</i> Roque & S.C.Ferreira	NE	NE	CR*
	<i>Chaptalia chapadensis</i> D.J.N.Hind	CR	CR	CR
	<i>Chionolaena jeffreyi</i> H.Rob.	NE	NE	NE
	<i>Dasyphyllum diamantinense</i> Saavedra & M.Monge	NE	VU	VU
	<i>Eremanthus harleyi</i> H.Rob.	NE	NE	NE
	<i>Eremanthus hatschbachii</i> H.Rob.	NE	EN	EN
	<i>Eremanthus leucodendron</i> Mattf.	EN	EN	EN
	<i>Fulcaldea stuessyi</i> Roque & V.A.Funk	NE	VU	VU
	<i>Hieracium stannardii</i> D.J.N.Hind	NE	VU	VU
	<i>Hoehnephytum almasensis</i> D.J.N.Hind	EN	EN	EN
	<i>Lapidia apicifolia</i> Roque & S.C.Ferreira	CR	VU	CR
	<i>Lasiolaena blanchetii</i> (Sch.Bip. ex Baker) R.M.King & H.Rob.	NE	VU	VU
	<i>Lasiolaena carvalhoi</i> D.J.N.Hind	NE	EN	EN
	<i>Lasiolaena duartei</i> R.M.King & H.Rob.	NE	NE	NE
	<i>Lasiolaena lychnophorioides</i> Roque, S.C.Ferreira & H.Rob.	NE	EN	EN
	<i>Lasiolaena pereirae</i> R.M.King & H.Rob.	NE	NE	NE
	<i>Lasiolaena santosii</i> R.M.King & H.Rob.	NE	VU	VU
	<i>Lepidaploa almasensis</i> (D.J.N.Hind) H.Rob.	NE	CR	CR
Asteraceae	<i>Lepidaploa bahiana</i> H.Rob.	NE	VU	VU
	<i>Lepidaploa tombadorensis</i> (H.Rob.) H.Rob.	NE	NE	NE
	<i>Lessingianthus carvalhoi</i> (H.Rob.) H.Rob.	NE	NE	NE
	<i>Lychnophora crispa</i> Mattf.	EN	NE	EN
	<i>Lychnophora harleyi</i> H.Rob.	NE	EN	EN
	<i>Lychnophora jeffreyi</i> H.Rob.	NE	VU	VU
	<i>Lychnophora sericea</i> D.J.N.Hind	CR	CR	CR
	<i>Lychnophora morii</i> H.Rob.	NE	EN	EN
	<i>Lychnophora santosii</i> H.Rob.	NE	EN	EN
	<i>Lychnophora spiciformis</i> Loeuille & Siniscalchi	CR	NE	CR
	<i>Mikania glandulosissima</i> W.C.Holmes & D.J.N.Hind	NE	NE	NE
	<i>Mikania grazielae</i> R.M.King & H.Rob.	NE	NE	NE
	<i>Mikania nelsonii</i> D.J.N.Hind	NE	NE	NE
	<i>Morithamnus crassus</i> R.M.King, H. Rob. & G.M.Barroso	NE	EN	EN
	<i>Paralychnophora atkinsae</i> D.J.N.Hind	EN	EN	EN
	<i>Paralychnophora harleyi</i> (H.Rob.) D.J.N.Hind	VU	VU	VU
	<i>Paralychnophora patriciana</i> D.J.N.Hind	EN	EN	EN
	<i>Porophyllum bahiense</i> D.J.N.Hind	VU	VU	VU
	<i>Pseudostiffia kingii</i> H.Rob.	NE	EN	EN
	<i>Scherya bahiensis</i> R.M.King & H.Rob.	NE	CR	CR
	<i>Semiria viscosa</i> D.J.N.Hind	NE	EN	EN

Asteraceae	<i>Senecio almasensis</i> Mattf.	CR	CR	CR
	<i>Senecio harleyi</i> D.J.N.Hind	NE	VU	VU
	<i>Senecio regis</i> H.Rob.	NE	NE	NE
	<i>Stenophalium eriodes</i> (Mattf.) Anderb.	VU	VU	VU
	<i>Stylotrichium corymbosum</i> (DC.) Mattf	NE	EN	EN
	<i>Stylotrichium edmundoi</i> G.M.Barroso	NE	EN	EN
	<i>Stylotrichium hortensiae</i> V.Amorim & Roque	NE	VU	VU
	<i>Stylotrichium glomeratum</i> Bautista, Rodr.Oubina & S.Ortiz	NE	CR	CR
	<i>Stylotrichium rotundifolium</i> Mattf.	NE	EN	EN
	<i>Stylotrichium sucrei</i> R.M.King & H.Rob.	NE	EN	EN
	<i>Trichogonia tombadorensis</i> R.M.King & H.Rob.	NE	EN	EN
	<i>Trichogoniopsis morii</i> R.M.King & H.Rob.	NE	CR	CR
	<i>Verbesina luetzelburgii</i> Mattf.	NE	EN	EN
	<i>Verbesina baccharifolia</i> Mattf.	NE	EN	EN
	<i>Vernonanthura fagifolia</i> (Gardner) H.Rob.	VU	VU	VU
	<i>Vernonia almasensis</i> D.J.N.Hind	NE	NE	NE
	<i>Vernonia leucodendron</i> (Mattf.) Macleish	NE	NE	NE
Bignoniaceae	<i>Handroanthus diamantinensis</i> Espírito-Santo & M.M.Silva-Castro	NE	NE	NE
	<i>Handroanthus grandiflorus</i> Espírito-Santo & M.M.Silva-Castro	NE	CR	CR
	<i>Jacaranda heterophylla</i> M.M.Silva-Castro	NE	NE	NE
Bromeliaceae	<i>Aechmea emmerichiae</i> Leme	NE	NE	NE
	<i>Cryptanthus arelii</i> H.Luther	NE	NE	NE
	<i>Cryptanthus diamantinensis</i> Leme.	NE	NE	NE
	<i>Dyckia brachystachya</i> Rauh & E.Gross	NE	NE	NE
	<i>Dyckia burlemarxii</i> L.B.Sm. & Read	NE	NE	NE
	<i>Dyckia hohenbergioides</i> Leme & Esteves	NE	NE	NE
	<i>Hohenbergia edmundoi</i> L.B.Sm. & Read	NE	NE	NE
	<i>Hohenbergia igatuensis</i> Leme	NE	NE	NE
	<i>Hohenbergia magnispina</i> Leme	NE	NE	NE
	<i>Hohenbergia pennae</i> E.Pereira	NE	NE	NE
	<i>Hohenbergia undulatifolia</i> Leme & H.Luther	NE	NE	NE
	<i>Neoregelia mucugensis</i> Leme	NE	NE	NE
	<i>Sincoraea albopicta</i> (Philcox) Louzada & Wand.	NE	EN	EN
	<i>Sincoraea amoena</i> Ule	EN	EN	EN
	<i>Sincoraea braunii</i> (Leme)	NE	NE	NE
	<i>Sincoraea burle-marxii</i> (L.B.Sm. & Read) Louzada & Wand.	NE	NE	NE
	<i>Sincoraea harleyi</i> (Leme & M.Machado) Louzada & Wand.	NE	EN	EN
	<i>Sincoraea hatschbachii</i> (Leme) Louzada & Wand.	NE	EN	EN
	<i>Sincoraea heleniceae</i> (Leme) Louzada & Wand.	NE	NE	NE
	<i>Sincoraea mucugense</i> (Wand. & A.A.Conc.) Louzada & Wand.	NE	VU	VU
<i>Sincoraea navioides</i> (L.B.Sm.) Louzada & Wand.	NE	NE	NE	

Bromeliaceae	<i>Sincoraea ophiuroides</i> (Louzada & Wand.) Louzada & Wand.	NE	VU	VU
	<i>Sincoraea riocontense</i> (Leme) Louzada & Wand.	NE	EN	EN
	<i>Vriesea fabioi</i> Leme	NE	NE	NE
Cactaceae	<i>Arrojadoa bahiensis</i> (P.J.Braun & Esteves) N.P.Taylor & Eggl	EN	EN	EN
	<i>Melocactus glaucescens</i> Buining & Brederoo	EN	EN	EN
	<i>Melocactus pachyacanthus</i> Buining & Brederoo	EN	EN	EN
	<i>Melocactus paucispinus</i> Heimen & R.J.Paul	VU	VU	VU
	<i>Micranthocereus hofackerianus</i> (P.J.Braun & Esteves) M.Machado	NE	NE	NE
	<i>Micranthocereus polyanthus</i> (Werderm.) Backeb.	EN	EN	EN
	<i>Micranthocereus streckeri</i> Van Heek & Van Criek.	CR	CR	CR
	<i>Pilosocereus glaucochrous</i> (Wenderm.) Byles & Rowley.	VU	VU	VU
Celastraceae	<i>Stephanocereus luetzelburgii</i> (Vaupel) N.P.Taylor & Eggl	NE	NE	NE
	<i>Elachyptera coriacea</i> Lombardi	DD	NE	DD
Combretaceae	<i>Monteverdia mucugensis</i> (R.M. Carvalho-Okano ex Biral & Groppo) Biral	NE	NE	NE
	<i>Pouteria subsessilifolia</i> Cronquist	NE	NE	NE
	<i>Evolvulus delicatus</i> C.V.Silva & Sim.-Bianch.	NE	NE	NE
	<i>Evolvulus harleyi</i> C.V.Silva & Sim.-Bianch.	NE	NE	NE
	<i>Ipomoea anamariae</i> L.V.Vasconcelos & Sim.-Bianch.	VU	EN	EN
	<i>Ipomoea serrana</i> Sim.-Bianch. & L.V.Vasconcelos	VU	EN	EN
	<i>Jacquemontia grisea</i> Buril	NE	NE	NE
Cucurbitaceae	<i>Jacquemontia robertsoniana</i> Buril & Sim.-Bianch.	NE	VU	VU
	<i>Jacquemontia staplesii</i> Buril	NE	NE	NE
	<i>Apodanthera villosa</i> C.Jeffrey	VU	VU	VU
Cyperaceae	<i>Cyperus almensis</i> D.A.Simpson	EN	EN	EM
	<i>Eleocharis morroi</i> D.A.Simpson	EN	NE	EM
	<i>Rhynchospora almensis</i> D.A.Simpson	NE	NE	NE
Droseraceae	<i>Drosera riparia</i> Rivadavia & Gonella	NE	NE	NE
Eriocaulaceae	<i>Actinocephalus herzogii</i> (Moldenke) Sano	EN	NE	EN
	<i>Comanthera bahiensis</i> (Moldenke) L.R.Parra & Giul	EN	EN	EN
	<i>Comanthera borbae</i> A.C.S.Pereira & Giul.	VU	NE	VU
	<i>Comanthera curralensis</i> (Moldenke) L.R.Parra & Giul.,	VU	VU	VU
	<i>Comanthera harleyi</i> (Moldenke) L.R.Parra & Giul.	VU	VU	VU
	<i>Comanthera hatschbachii</i> (Moldenke) L.R.Parra & Giul.	VU	NE	VU
	<i>Comanthera mucugensis</i> (Giul.) L.R.Parra & Giul.	EN	EN	EN
	<i>Leiothrix raymondii</i> Giul. & D.M.Silva	EN	NE	EN
	<i>Paepalanthus almasensis</i> Moldenke	EN	NE	EN
	<i>Paepalanthus barbulatus</i> Herzog	VU	NE	VU
	<i>Paepalanthus carvalhoi</i> Giul. & E.Miranda	NE	VU	VU
	<i>Paepalanthus cinereus</i> Giul. & L.R.Parra	EN	NE	EN
	<i>Paepalanthus contasensis</i> Moldenke	EN	NE	EN
<i>Paepalanthus harleyi</i> Moldenke	EN	NE	EN	

Eriocaulaceae	<i>Paepalanthus inopinatus</i> Moldenke	EN	NE	EN
	<i>Paepalanthus luetzelburgii</i> Herzog	EN	NE	EN
	<i>Paepalanthus multicapitatus</i> Giul. & E.Miranda	NE	NE	NE
	<i>Paepalanthus oblongifolius</i> Giul. & E.Miranda	NE	NE	NE
	<i>Paepalanthus umbrosus</i> Giul. & E.Miranda	NE	VU	VU
Euphorbiaceae	<i>Manihot longiracemosa</i> P.Carvalho & M.Martins,	NE	NE	NE
	<i>Manihot refexifolia</i> P.Carvalho & M.Martins	NE	NE	NE
	<i>Euphorbia teres</i> M.Machado & Hofacker	NE	NE	NE
	<i>Euphorbia apparicana</i> Rizzini	EN	NE	EN
Fabaceae	<i>Abarema diamantina</i> E.Guerra, Iganci & M.PMorim	NE	NE	NE
	<i>Aeschynomene chicocesariana</i> D.B.O.S.Cardoso & G.Ramos	NE	NE	CR*
	<i>Bauhinia funchiana</i> Vaz & G.P.Lewis	NE	CR	CR
	<i>Calliandra asplenioides</i> (Nees) Renvoize	NE	NE	NE
	<i>Calliandra bahiana</i> Renvoize	NE	NE	NE
	<i>Calliandra bromelioides</i> E.R.Souza & L.P.Queiroz	NE	NE	NE
	<i>Calliandra calycina</i> Benth.	NE	NE	NE
	<i>Calliandra coccinea</i> Renvoize	NE	NE	NE
	<i>Calliandra crassipes</i> Benth.	NE	NE	NE
	<i>Calliandra cumbucana</i> Renvoize	NE	NE	NE
	<i>Calliandra debilis</i> Renvoize	NE	EN	EN
	<i>Calliandra elegans</i> Renvoize	NE	NE	NE
	<i>Calliandra erubescens</i> Renvoize	NE	NE	NE
	<i>Calliandra fasciculata</i> Benth.	NE	NE	NE
	<i>Calliandra fuscipila</i> Harms	NE	NE	NE
	<i>Calliandra ganevii</i> Barneby	NE	VU	VU
	<i>Calliandra geraensis</i> E.R.Souza & L.P.Queiroz	NE	VU	VU
	<i>Calliandra germana</i> Barneby	NE	NE	NE
	<i>Calliandra hirsuticaulis</i> Harms	NE	NE	NE
	<i>Calliandra hirtiflora</i> Benth.	NE	NE	NE
	<i>Calliandra hygrophila</i> Mackinder & G.P.Lewis	NE	NE	NE
	<i>Calliandra imbricata</i> E.R.Souza & L.P.Queiroz	DD	NE	DD
	<i>Calliandra involuta</i> Mackinder & G.P.Lewis	NE	VU	VU
	<i>Calliandra lanata</i> Benth.	NE	NE	NE
	<i>Calliandra leptopoda</i> Benth.	NE	NE	NE
	<i>Calliandra lewisii</i> E.R.Souza & L.P.Queiroz	NE	NE	NE
	<i>Calliandra lintea</i> Barneby	NE	NE	NE
	<i>Calliandra longipinna</i> Benth.	NE	NE	NE
	<i>Calliandra luetzelburgii</i> Harms	NE	NE	NE
	<i>Calliandra mucugeana</i> Renvoize.	NE	NE	NE
	<i>Calliandra nebulosa</i> Barneby	NE	NE	NE
	<i>Calliandra paganuccii</i> E.R.Souza	NE	EN	EN

Fabaceae	<i>Calliandra paterna</i> Barneby	NE	NE	NE
	<i>Calliandra pilgerana</i> Harms	NE	NE	NE
	<i>Calliandra pubens</i> Renvoize	NE	VU	VU
	<i>Calliandra renvoizeana</i> Barneby	NE	NE	NE
	<i>Calliandra semiseputa</i> Barneby	VU	VU	VU
	<i>Calliandra sincorana</i> Harms	NE	NE	NE
	<i>Calliandra stelligera</i> Barneby	NE	VU	VU
	<i>Calliandra viscidula</i> Benth.	NE	NE	NE
	<i>Chamaecrista anamariae</i> Conc., L.P.Queiroz & G.P.Lewis	EN	EN	EN
	<i>Chamaecrista arboae</i> Barneby	NE	EN	EN
	<i>Chamaecrista axilliflora</i> H.S.Irwin & Barneby	NE	NE	NE
	<i>Chamaecrista botryoides</i> Conc., L.P.Queiroz & G.P.Lewis	NE	END	EN
	<i>Chamaecrista catolesensis</i> Conc., L.P.Queiroz & G.P.Lewis	VU	VU	VU
	<i>Chamaecrista depauperata</i> Conc., L.P.Queiroz & G.P.Lewis	NE	EN	EN
	<i>Chamaecrista punctulifera</i> (Harms) H.S.Irwin & Barneby	NE	VU	VU
	<i>Chamaecrista sincorana</i> (Harms) H.S.Irwin & Barneby.	NE	VU	VU
	<i>Harpalyce lanata</i> L.P.Queiroz	EN	EN	EN
	<i>Harpalyce riparia</i> São-Mateus, L.P.Queiroz & D.B.O.S.Cardoso	NE	NE	EN*
	<i>Luetzelburgia harleyi</i> D.Cardoso, L.P.Queiroz & H.C.Lima	EN	EN	EN
	<i>Luetzelburgia neurocarpa</i> D.Cardoso, L.P.Queiroz & H.C.Lima	CR	NE	CR
<i>Mimosa mensicola</i> Barneby.	EN	EN	EN	
<i>Mimosa subenervis</i> Benth.	NE	NE	NE	
<i>Mimosa crumenarioides</i> L.P.Queiroz & G.P.Lewis	NE	NE	NE	
<i>Poiretia bahiana</i> C.Müller	LC	NE	LC	
Gentianaceae	<i>Macrocarpaea illecebrosa</i> J.R.Grant	NE	VU	VU
	<i>Schultesia bahiensis</i> E.F.Guim. & Fontella	LC	NE	LC
Gesneriaceae	<i>Sinningia harleyi</i> Wiehler & Chautems	EN	EN	EN
Iridaceae	<i>Pseudiris speciosa</i> Chuckr & Giul.	NE	NE	NE
	<i>Trimezia sincorana</i> Ravenna.	NE	NE	NE
Lamiaceae	<i>Cyanocephalus delicatulus</i> (Harley) Harley & J.FB.Pastore	EN	EN	EN
	<i>Eplingiella cuniloides</i> (Epling) Harley & J.FB.Pastore	VU	END	VU
	<i>Eriope anamariae</i> Harley	EN	EN	EN
	<i>Eriope confusa</i> Harley	EN	EN	EN
	<i>Eriope crassifolia</i> Mart. ex Benth.	NE	NE	NE
	<i>Eriope exaltata</i> Harley	NE	NE	NE
	<i>Eriope ganevii</i> Harley	NE	NE	NE
	<i>Eriope glandulosa</i> (Harley) Harley	NE	NE	NE
	<i>Eriope luetzelburgii</i> Harley	VU	VU	VU
	<i>Eriope montana</i> Harley	VU	VU	VU
	<i>Eriope obovata</i> Epl.	NE	EN	EN
	<i>Eriope polyphylla</i> Mart. ex Benth.	NE	VU	VU

Lamiaceae	<i>Eriope sincorana</i> Harley	NT	NE	NT
	<i>Eriope viscosa</i> Harley & Walsingham	NE	EN	EN
	<i>Hyptis bahiensis</i> Harley	EN	EN	EN
	<i>Hyptis ganevii</i> Harley	NE	NE	NE
	<i>Leptohyptis pinheiroi</i> (Harley) Harley & J.F.B.Pastore	EN	EN	EM
	<i>Medusantha carvalhoi</i> (Harley) Harley & J.F.B.Pastore	VU	NE	VU
	<i>Mesosphaerum irwinii</i> (Harley) Harley & J.F.B.Pastore	NT	NE	NT
	<i>Oocephalus argyrophyllus</i> (Harley) Harley & J.F.B.Pastore	EN	NE	END
	<i>Oocephalus ganevii</i> Harley	NE	VU	VU
	<i>Oocephalus hagei</i> (Harley) Harley & J.F.B.Pastore Harley	VU	NE	VU
	<i>Oocephalus halimifolius</i> (Mart. ex Benth.) Harley & J.F.B.Pastore	EN	EN	EN
	<i>Oocephalus nubicola</i> (Harley) Harley & J.F.B.Pastore	EN	CR	CR
	<i>Oocephalus pauciflorus</i> (Harley) Harley & J.F.B.Pastore	NE	END	END
	<i>Oocephalus rigens</i> Harley	NE	VU	VU
	<i>Oocephalus rhodocalyx</i> A.Souares & Harley	NE	NE	CR*
	<i>Oocephalus silvinae</i> (Harley) Harley & J.F.B.Pastore	EN	EN	EN
	<i>Oocephalus tenuithyrus</i> Harley	NE	EN	EN
Lauraceae	<i>Aniba subbullata</i> H.L.Ribeiro & P.L.R.Moraes	NE	NE	NE
	<i>Ocotea vegrans</i> P.L.R.Moraes & van der Werff	NE	EN	EN
	<i>Ocotea rohweri</i> P.L.R.Moraes & van der Werff	NE	VU	VU
Lentibulariaceae	<i>Genlisea exhibitionista</i> Rivadavia & A.Fleischm.	NE	EN	EN
	<i>Genlisea uncinata</i> P.Taylor & Fromm	NE	EN	EN
	<i>Utricularia catolesensis</i> G.L.Campos, Cheek & Giul.	NE	NE	NE
Loganiaceae	<i>Spigelia cremnophila</i> Zappi & Lucas	NE	EN	EN
	<i>Spigelia flava</i> Zappi & Harley	NE	VU	VU
Loranthaceae	<i>Ligaria teretiflora</i> (Rizzini) Kuijt	NE	EN	EN
Lythraceae	<i>Cuphea sincorana</i> T.B.Cavalc.	DD	NE	DD
	<i>Diplusodon argyrophyllus</i> T.B.Cavalc.	CR	CR	CR
	<i>Diplusodon bahiensis</i> T.B.Cavalc.	NE	VU	VU
	<i>Diplusodon parvifolius</i> Mart. ex DC.	NT	NE	NT
Malpighiaceae	<i>Camarea elongata</i> Mamede	NE	VU	VU
	<i>Diplopterys bahiana</i> W.R.Anderson & C.Davis	NE	CR	CR
Malvaceae	<i>Ayenia noblickii</i> Cristóbal	NE	CR	CR
	<i>Gaya dentata</i> Krapov.	NE	EN	EN
	<i>Pavonia almasana</i> Ulbr.	EM	EN	EN
	<i>Pavonia palmeirensis</i> Krapov.	NE	CR	CR
	<i>Helicteres rufipila</i> Cristóbal	NE	CR	CR
	<i>Rayleya bahiensis</i> Cristobal.	NE	CR	CR
Melastomataceae	<i>Cambessedesia gracilis</i> Wurdack	EN	EN	EN
	<i>Cambessedesia hermogenesii</i> A.B.Martins	EN	EN	EN
	<i>Cambessedesia rupestris</i> A.B.Martins	NE	EN	EN

	<i>Chaetostoma luetzelburgii</i> Marckgr.	NE	NE	NE
	<i>Chaetostoma parvulum</i> Marckgr.	NE	NE	NE
	<i>Lavoisiera harleyi</i> Wurdack	EN	EN	EN
	<i>Marcetia alba</i> Ule	CR	CR	CR
	<i>Marcetia bahiana</i> (Ule) A.B.Martins	EN	EN	EN
	<i>Marcetia candolleana</i> A.K.A.Santos & A.B.Martins	NE	VU	VU
	<i>Marcetia eimeariana</i> A.B.Martins & Woodgyer	NE	VU	VU
	<i>Marcetia formosa</i> Wurdack	EN	EN	EN
	<i>Marcetia grandiflora</i> Wurdack	NE	EN	EM
	<i>Marcetia harleyi</i> Wurdack	NE	NE	NE
	<i>Marcetia lanuginosa</i> Wurdack	NE	NE	NE
	<i>Marcetia luetzelburgii</i> Markgr.	EN	EN	EN
	<i>Marcetia lychnophoroides</i> A.B.Martins	EN	EN	EN
	<i>Marcetia macrophylla</i> Wurdack	NE	NE	NE
	<i>Marcetia mucugensis</i> Wurdack	NE	NE	NE
	<i>Marcetia nervulosa</i> Markgr.	NE	VU	VU
	<i>Marcetia nummularia</i> Markgr.	EN	EN	EN
	<i>Marcetia oxycoccoides</i> Wurdack & A.B.Martins	EN	EN	EN
	<i>Marcetia sincorensis</i> Wurdack	NE	VU	VU
	<i>Marcetia velutina</i> Markgr.	NE	NE	NE
Melastomataceae	<i>Marcetia viscida</i> Wurdack	VU	VU	VU
	<i>Microlicia amblysepala</i> Ule	NE	VU	VU
	<i>Microlicia aurea</i> Wurdack	NE	VU	VU
	<i>Microlicia balsamifera</i> (DC.) Mart.	NE	VU	VU
	<i>Microlicia blanchetiana</i> Cogn.	NE	VU	VU
	<i>Microlicia carrasci</i> Markgr.	NE	NE	NE
	<i>Microlicia catolensis</i> Woodgyer & Zappi	NE	EN	EN
	<i>Microlicia chrysantha</i> Wurdack	NE	NE	NE
	<i>Microlicia comparilis</i> Wurdack	NE	NE	NE
	<i>Microlicia contasensis</i> Woodgyer & Zappi	NE	NE	NE
	<i>Microlicia flavovirens</i> Woodgyer & Zappi	NE	VU	VU
	<i>Microlicia giuliettiana</i> A.B.Martins & Almeda	NE	NE	NE
	<i>Microlicia harleyi</i> Wurdack	NE	EN	EN
	<i>Microlicia hatschbachii</i> Wurdack	NE	NE	NE
	<i>Microlicia hirta</i> Pataro & R. Romero	NE	NE	VU*
	<i>Microlicia intercalycina</i> Pataro & R. Romero	NE	NE	EN*
	<i>Microlicia isostemon</i> Wurdack	NE	NE	NE
	<i>Microlicia leucopetala</i> Wurdack	NE	EN	EN
	<i>Microlicia luetzelburgii</i> Markgr.	NE	NE	NE
	<i>Microlicia lutea</i> Markgr.	NE	NE	NE
	<i>Microlicia macropetala</i> Pataro & R. Romero	NE	NE	EN*

Melastomataceae	<i>Microlicia minima</i> Markgr.	NE	NE	NE
	<i>Microlicia monticola</i> Wurdack	NE	NE	NE
	<i>Microlicia morii</i> Wurdack	NE	EN	EN
	<i>Microlicia mucugensis</i> (Wurdack) Almeda & A.B.Martins	NE	EN	EN
	<i>Microlicia noblickii</i> (Wurdack) A.B.Martins & Almeda	NE	EN	EN
	<i>Microlicia oligochaeta</i> Wurdack	NE	EN	EN
	<i>Microlicia petasensis</i> Wurdack	NE	VU	VU
	<i>Microlicia pinheiroi</i> Wurdack	NE	EN	EN
	<i>Microlicia plumosa</i> Woodgyer & Zappi	NE	VU	VU
	<i>Microlicia pulchra</i> Pataro & R.Romero	NE	NE	EN*
	<i>Microlicia sincorensis</i> (DC.) Mart.	NE	VU	VU
	<i>Microlicia subaequalis</i> Wurdack	NE	EN	EN
	<i>Microlicia subalata</i> Wurdack	NE	CR	CR
	<i>Microlicia taxifolia</i> Naudin	NE	NE	NE
	<i>Microlicia torrendi</i> Brade.	NE	NE	NE
	<i>Microlicia wurdackiana</i> Almeda & A.B.Martins	NE	NE	NE
	<i>Rupestrea carvalhoana</i> (Baumgratz & Souza) Almeda, Michelang. & R.Goldenb.	CR	NE	CR
	<i>Rupestrea johnwurdackiana</i> (Baumgratz & Souza) Michelang., Almeda & R.Goldenb.	EN	EN	EN
	<i>Pleroma bracteolatum</i> (J.G.Freitas, A.K.A.Santos & R.P.Oliveira) P.J.F.Guim. & Michelang.	NE	NE	NE
	<i>Pleroma rubrum</i> J.G.Freitas	EN	NE	EN
	<i>Pterolepis parnassiifolia</i> (DC.) Triana	NE	NE	NE
	<i>Pterolepis rotundifolia</i> Wurdack	NE	EN	EN
	<i>Tibouchina barnebyana</i> Wurdack	NE	NE	NE
	<i>Tibouchina carvalhoi</i> Wurdack	NE	VU	VU
	<i>Tibouchina comosa</i> J. G. Freitas, A. K. A. Santos & R. P.Oliveira	NE	NE	NE
	<i>Tibouchina luetzelburgii</i> Markgr.	NE	EN	EN
<i>Tibouchina oreophila</i> Wurdack	NE	NE	NE	
<i>Tibouchina pereirae</i> Brade & Markgr.	NE	NE	NE	
Moraceae	<i>Sorocea ganevii</i> Castro & Rapini	NE	NE	NE
Myrtaceae	<i>Eugenia mucugensis</i> Sobral	NE	NE	NE
	<i>Marlierea jacobinensis</i> (Berg) Mattos	NE	NE	NE
	<i>Myrcia almasensis</i> NicLugh.	NE	EN	EN
	<i>Myrcia jacobinensis</i> Mattos	NE	NE	NE
	<i>Myrcia lucasiae</i> R.B.Almeida, Antar & B.S.Amorim	NE	NE	NE
	<i>Myrcia lughadhai</i> B.S.Amorim	NE	VU	VU
	<i>Myrcia pseudovenulosa</i> Stadnik & Sobral	NE	EN	EN
Ochnaceae	<i>Sauvagesia insignis</i> (Ule) Sastre	NE	EN	EN
	<i>Sauvagesia nitida</i> Zappi & E.Lucas	NE	NE	NE
	<i>Sauvagesia oliveirae</i> Harley & Giul.	NE	NE	NE
	<i>Sauvagesia paniculata</i> Cardoso & A.A.Conceição	VU	CR	CR

Ochnaceae	<i>Sauvagesia ribeiroi</i> Harley & Giul.	NE	NE	NE
	<i>Sauvagesia semicylindrifolia</i> Sastre.	NE	NE	NE
Orchidaceae	<i>Adamantina miltonioides</i> Van den Berg & C.N.Gonç.	CR	CR	CR
	<i>Bulbophyllum seabrense</i> Campacci	NE	NE	NE
	<i>Cattleya elongata</i> Barb. Rodr.	NE	NE	NE
	<i>Cattleya luetzelburgii</i> Van den Berg	NE	NE	NE
	<i>Cattleya pfisteri</i> (Pabst & Senghas) Van den Berg	NE	NE	NE
	<i>Cattleya sincorana</i> (Schltr.) Van den Berg	EN	NE	EN
	<i>Cattleya tenuis</i> Campacci & Vedovello	EN	EM	EN
	<i>Encyclia joaosaiana</i> Campacci	NE	NE	NE
	<i>Gomesa adamantina</i> (Marçal & Cath.) M.W.Chase & N.H.Williams	NE	NE	NE
	<i>Gomesa sincorana</i> (Campacci & Cath.) M.W.Chase & N.H.Williams	NE	VU	VU
	<i>Habenaria pseudohamata</i> Toscano	NE	NE	NE
	<i>Prescottia mucugensis</i> C.O.Azevedo & Van den Berg	NE	NE	NE
	<i>Prosthechea moojenii</i> (Pabst) W.E. Higgins	NE	NE	NE
	<i>Sarcoglottis riocontensis</i> E.C.Smidt & Toscano	NE	NE	DD*
	<i>Thelyschista ghillanyi</i> (Pabst) Garay	VU	NE	VU
	<i>Veyretia sincorensis</i> (Schltr.) Szlach.	NE	EN	EN
Orobanchaceae	<i>Physocalyx scaberrimus</i> Philcox	NE	NE	NE
Passifloraceae	<i>Passiflora mucugeana</i> T.S.Nunes & L.P.Queiroz	NE	EN	EN
	<i>Piriqueta abairana</i> Arbo	NE	EN	EN
	<i>Piriqueta asperifolia</i> Arbo	NE	VU	VU
	<i>Piriqueta carnea</i> Urb.	NE	NE	NE
	<i>Piriqueta crenata</i> L.Rocha, I.M.Souza & Arbo	NE	NE	CR*
	<i>Piriqueta dentata</i> Arbo	NE	NE	NE
	<i>Piriqueta douradinha</i> Arbo	NE	EN	EN
	<i>Piriqueta flammea</i> (Suesseng.) Arbo	NE	CR	CR
	<i>Piriqueta nanuzae</i> Arbo	NE	EN	EN
	<i>Piriqueta revoluta</i> Arbo	NE	NE	NE
	<i>Turnera asymmetrica</i> Arbo	NE	NE	NE
	<i>Turnera caatingana</i> Arbo	NE	EN	EN
	<i>Turnera harleyi</i> Arbo	NE	VU	VU
	<i>Turnera involucrata</i> Arbo	NE	NE	NE
	<i>Turnera luetzelburgii</i> Sleumer	NE	NE	NE
	<i>Turnera stenophylla</i> Urb.	NE	EN	EN
Phyllanthaceae	<i>Phyllanthus gongyloides</i> Cordeiro & Carn.-Torres	NE	EN	EN
	<i>Phyllanthus sincorensis</i> G.L.Webster.	NE	NE	NE
	<i>Phyllanthus spartioides</i> Pax & K.Hoffm.	NE	NE	NE
Plantaginaceae	<i>Philcoxia bahiensis</i> V.C.Souza & Harley	NE	CR	CR
	<i>Philcoxia tuberosa</i> M.L.S.Carvalho & L.P.Queiroz	CR	CR	CR
	<i>Stemodia harleyi</i> B.L.Turner	VU	VU	VU

Poaceae	<i>Abildgaardia disticha</i> Lye	NE	NE	NE
	<i>Abildgaardia papilosa</i> Kral & M.Strong	NE	NE	NE
	<i>Andropogon durifolium</i> Renvoize	NE	NE	NE
	<i>Axonopus grandifolius</i> Renvoize	VU	NE	VU
	<i>Axonopus tenuis</i> Renvoize	NE	NE	NE
	<i>Dichantherium arenicola</i> A.O.Matos & R.P.Oliveira	NE	NE	NE
	<i>Dichantherium assurgens</i> (Renvoize) Zuloaga	NE	NE	NE
	<i>Dichantherium cumbucana</i> (Renvoize) Zuloaga	NE	VU	VU
	<i>Merostachys ramosissima</i> Send.	NE	NE	NE
	<i>Otachyrium aquaticum</i> Send. & Soderstr.	NE	NE	NE
	<i>Panicum animarum</i> Renvoize	NE	NE	NE
	<i>Panicum belmonte</i> Renvoize	NE	NE	NE
	<i>Panicum noterophilum</i> Renvoize	NE	NE	NE
	<i>Streptostachys lanciflora</i> R.P.Oliveira & Longhi-Wagner	DD	NE	DD
	<i>Urochloa decidua</i> Zuloaga & Morrone	NE	NE	NE
Polygalaceae	<i>Polygala sincorense</i> Chodat.	NE	NE	NE
	<i>Polygala trifurcata</i> Chodat	EN	EN	EN
	<i>Polygala tuberculata</i> Chodat	NE	NE	NE
Portulacaceae	<i>Portulaca werdermanni</i> Poelln.	NE	NE	NE
Pteridaceae	<i>Doryopteris trilobata</i> J.Prado	EN	NE	EN
Rubiaceae	<i>Borreria bahiana</i> E.L.Cabral	NE	NE	NE
Santalaceae	<i>Phoradendron harleyi</i> Kuijt	NE	NE	NE
Velloziaceae	<i>Barbacenia contasana</i> L.B.Sm. & Ayensu	EN	EN	EN
	<i>Barbacenia regis</i> L.B.Sm.	EN	EN	EN
	<i>Vellozia burlemarxii</i> L.B.Sm. & Ayensu	NE	NE	NE
	<i>Vellozia canelinha</i> Mello-Silva	CR	NE	CR
	<i>Vellozia joly</i> L.B. Smith & Ayensu	NE	NE	NE
	<i>Vellozia pyrantha</i> A.A.Conceição	NE	NE	NE
	<i>Vellozia punctulata</i> Seub.	NE	NE	NE
	<i>Vellozia sincorana</i> L.B.Sm. & Ayensu.	NE	NE	NE
Verbenaceae	<i>Lippia alnifolia</i> Schauer	VU	VU	VU
	<i>Lippia insignis</i> Moldenke	VU	NE	VU
	<i>Lippia morii</i> Moldenke	EN	EN	EN
	<i>Lippia subracemosa</i> Mansf.	NE	NE	NE
	<i>Stachytarpheta almasensis</i> Mansf..	EN	EN	EN
	<i>Stachytarpheta arenaria</i> S.Atkins	NE	VU	VU
	<i>Stachytarpheta atkinsiae</i> Harley & Giul.	NE	NE	EN*
	<i>Stachytarpheta bromleyana</i> S. Atkins	NE	NE	NE
	<i>Stachytarpheta froesii</i> Moldenke	EN	EN	EN
	<i>Stachytarpheta galactea</i> S.Atkins	DD	NE	DD
	<i>Stachytarpheta ganevii</i> S.Atkins	NE	VU	VU



Verbenaceae	<i>Stachytarpheta guedesii</i> S. Atkins	NE	VU	VU
	<i>Stachytarpheta lacunosa</i> Mart. ex Schauer	VU	NE	VU
	<i>Stachytarpheta lychnitis</i> Mart. ex Schauer	VU	NE	VU
	<i>Stachytarpheta piranii</i> S. Atkins	NE	NE	NE
	<i>Stachytarpheta quadrangula</i> Nees & Mart.	NE	NE	NE
	<i>Stachytarpheta radlkoferiana</i> Mansf.	VU	VU	VU
	<i>Stachytarpheta tuberculata</i> S. Atkins	NE	VU	VU
Xyridaceae	<i>Xyris almae</i> Kral & Wand.	NT	NE	NT
	<i>Xyris fibrosa</i> Kral & Wand.	CR	CR	CR
	<i>Xyris mertesiana</i> Koern. ex Malme	EN	EN	EN
	<i>Xyris morii</i> Kral & L.B.Sm.	EN	EN	EN
	<i>Xyris phaeocephala</i> Kral & Wand.	EN	EN	EN
	<i>Xyris picea</i> Kral & Wand.	EN	EN	EN
	<i>Xyris sincorana</i> Kral & Wand.	EN	NE	EN